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Colonization and antifungals susceptibility patterns of Candida species isolated from hospitalized patients in ICUs and NICUs

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ABSTRACT

Background: Several studies have shown that there are an increasing in invasive candidiasis during 2-3 last decades. Although, *Candida albicans* is considered as the most common candidiasis agents, other non-albicans such as *C. glabrata*, *C. krusei*, *C. parapsilosis*, and *C. tropicalis* were raised as infectious agents. Resistance to fluconazole among non-albicans species is an important problem for clinicians during therapy and prophylaxis.

Objectives: The aim of current study was to detect the *Candida* species from hospitalized neonatal and children in intensive care units (ICUs) and neonatal intensive care units (NICUs). In addition, the susceptibility of isolated agents were also evaluated against three antifungals. *Materials and Methods:* In the present study 298 samples including 98 blood samples, 100 urines and 100 swabs from oral cavity were inoculated on CHROMagar Candida. Initial detection was done according to the coloration colonies on CHROMagar Candida. Morphology on cornmeal agar, germ tube formation and growth at 45°C were confirmed isolates. Amphotericin B, fluconazole and terbinafine (Lamisil) were used for the susceptibility tests using microdilution method.

Results: In the present study 21% and 34% of urines and swabs from oral cavity were positive for Candida species, respectively. The most common species was C. albicans (62.5%) followed by C. tropicalis (15.6%), C. glabrata (6.3%) and Candida species (15.6%). Our study indicated that the most tested species of Candida, 70.3% were sensitive to fluconazole at the concentration of $\leq 8 \, \mu g/mL$. Whereas 9 (14.1%) of isolates were resistant to amphotericine B at $\geq 8 \, \mu g/mL$.

Conclusions: This study demonstrates the importance of species identification and antifungals susceptibility testing for hospitalized patients in ICUs and NICUs wards.

Implication for health policy/practice/research/medical education:

The colonization of *Candida* species (especially, with high counts of *Candida*) and resistance to routine used antifungals among hospitalized patients in ICUs and NICUs can be a challenge for clinicians and patients.

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1. Background

Candida species are the commonest fungal infections agents and several studies showed that there are an increasing in candidiasis during the last several decades (1-3). In addition, the systemic candidiasis continues to be a major cause of morbidity and mortality among the compromised patients with a rate as high as 40%-60% (4). The presence of Candida species in urinary system (candiduria) is one of the most important challenges for patients, clinicians and laboratory workers. Candiduria is a common finding in patients with multiple predisposing factors such as, diabetes mellitus, indwelling urinary catheters, antimicrobials and corticosteroids users (5-9). In addition, the frequency of candiduria was more prevalent among the hospitalized patients in intensive care units (ICUs) and neonatal intensive care units (NICUs) (10-12). Physicians usually do not prescribed systemic antifungal therapy for the asymptomatic candiduria and have believe that disease benign and represent only colonization in urinary tract system. On the other hand, symptomatic candiduria may be associated with invasive candidiasis such as, candidemia (7,13). Colonization of the neonatal oral mucosal by Candida species is the first step for invasive candidiasis. Mendiratta et al (14) have believed that colonization by Candida species is more prevalent among preterms (33.9%) than terms (10%).

Although, Candida albicans is considered as the most common candidiasis agents, other non-albicans such as C. glabrata, C. krusei, C. parapsilosis, and C. tropicalis were increased during 2-3 last decades (5,15,16). Researchers have shown that non-albicans species have an important role in the increasing of mortality rate among the patients with invasive candidiasis due to inherently resistant to antifungals and increasing resistant due to prophylaxis (17-19). Systemic antifungals (amphotericine B, fluconazole, terbinafine, caspofungin) are usually prescribed for systemic mycosis, but different susceptibilities to etiologic agents were observed both in vivo and in vitro. Several studies have shown that resistance to antifungals increased during last decades, especially in teaching hospitals (20). Resistance to fluconazole among non-albicans is an important problem for clinicians during therapy and prophylaxis (21-23).

2. Objectives

The aim of current study was to detect *Candida* species from the hospitalized neonatal and children in NICUs and ICUs in Ahvaz. In addition, the susceptibility patterns of causative agents were evaluated against amphotericin B, fluconazole and

terbinafine antifungals.

3. Materials and Methods

3.1. Patients and sampling

In the present study 298 specimens including; 98 blood, 100 urines and 100 swabs from oral cavity were taken from infants and children hospitalized in NICUs and ICUs, Aboozar, Sina and Imam Khomeini hospitals, Ahvaz Jundishapur University of Medical Sciences. Two to 5 mL of venous blood were sampled and injected into blood culture media (Baharafshan, Iran). All cultured media were incubated at 37°C for 1 week and examined daily. After 3, 5 and 7 days, 10 µL of culture media were transferred to CHROMagar Candida (CHROMagar Candida, France) and incubated at 37°C for 2-3 days. All media were examined for colored colonies of Candida species. Swabs from oral cavity were inoculated on CHROMagar Candida and incubated at 37°C for 2-3 days. Cultured media were daily examined for fungal growth and discarded when no growth were seen. Ten microliter of urine samples were also spread on the surface of CHROMagar Candida and incubated at 37°C for 48-72 hours. Growth colonies were counted and the numbers of microorganisms per each milliliter of urine (CFU/mL) were calculated.

3.2. Identification of organisms

Initial detection was done according to the coloration colonies on CHROMagar Candida culture medium. *C. albicans, C. glabrata*, and *C. tropicalis* were produced green, pink and blue colored colonies, respectively. In addition morphology on cornmeal agar (HiMedia, India) supplemented with 1% Tween 80, germ tube formation and growth at 45°C were confirmed isolates. All isolated strains subcultured on slants of Sabouraud dextrose agar, SDA (Merck, Germany) and stored at room temperature until use.

3.3. Antifungal tests

In the present study, amphotericin B, fluconazole and terbinafine antifungals were used for the susceptibility tests using microdilution method according to Clinical and Laboratory Standards Institute (CLSI) protocol (24).

3.4. Ethical issues

The research followed the tenets of the Declaration of Helsinki. Informed consents were obtained. All patients took part in this study voluntary. The research was approved by ethical committee of Ahvaz Jundishapur University of Medical Sciences (ethic code: ajums.REC.1392.172).

3.5. Statistical analysis

In the present study the frequency of candiduria and theirs causative agents among both case and control were calculated and tabulated. All data were analyzed by SPSS 15 (SPSS Inc, Chicago, IL, USA). Values were shown as the percent frequency.

4. Results

4.1. Results of isolated organisms

In the present study, 34% of swabs from oral cavity were positive for *Candida* species including; 15 (44.1%) cases males and 19 (55.9%) cases females. As Table 1 shows, the 38.2% of positive cases were included at the rage less than 1 week, whereas only 1 case was observed at 7-12 years old. The most common species was *C. albicans* 24 (61.5%) followed by *Candida* species 7 (18%), *C. glabrata* 5 (12.9%), and *C. tropicalis* 3 (7.6%). Our study shows that 18 (46.1%) of samples yielded more than 50 colonies per each swab (Table 2). In our study in 5 cases, 2 different *Candida* species were isolated.

Our study shows that 21% of urines samples were yielded *Candida* species, including 12 (57.1%) males and 9 (42.9%) females (Table 3). Totally 85.6% of cases have less than 6 months year old. The most common agents was *C. albicans* 16 (64%) followed by *C. glabrata* 5 (20%), *Candida* species 3 (12%), and *C. tropicalis* 1 (4%). Polymicrobial growth in urine cultures was identified in three cases. Table 4 illustrates the details about colony counts of urines cultures. Colony counts more than 10000 CFU/mL was detected at

Table 1. Age range and sex sampled cases with positive culture (swabs)

Age range	Male	Female	Total
<1 week	7 (20.6%)	6 (17.6%)	13 (38.2%)
1-4 weeks	2 (5.9%)	2 (5.9%)	4 (11.8%)
1-6 months	2 (5.9%)	1 (3.0%)	3 (8.9%)
7-12 months	1 (3.0%)	0 (0.0%)	1 (3.0%)
1-6 years	2 (5.9%)	7 (20.6%)	9 (26.5%)
7-12 years	1 (3.0%)	3 (8.8%)	4 (11.8%)
Total	15 (44.1%)	19 (55.9%)	34 (100%)

9 (36%) cases of candiduria. Totally, in the present study the most common isolate was *C. albicans* (40, 62.5%), followed by *C. glabrata* (10, 15.6%), *C. tropicalis* (4, 6.3%) and *Candida* species (10, 15.6%).

In the present study all blood cultures were negative for fungal growth.

4.2. Results of antifungals

In the present study 64 isolates of *Candida* species including *C. albicans* (40 isolates), *C. glabrata* (10 isolates), *C. tropicalis* (4 isolates) and *Candida* species (10 isolates) were examined against 3 systemic antifungal drugs (amphotericine B, fluconazole and terbinafine). Amphotericine B were inhibited the growth of tested isolates at the rage of $0.5 - \ge 8 \, \mu \text{g/mL}$. The most of the isolates (32.8%) had MIC=4 $\, \mu \text{g/mL}$, whereas MIC for only 9 (14.1%) of isolates was more than 8 $\, \mu \text{g/mL}$ (Table 5).

The range of MICs to fluconazole of the 64 isolates was from 0.03125 to 64 µg/mL (Table 6). In addition the MIC 50 and MIC 90 of these isolates were 0.25 and 0.0625 µg/mL, respectively. The susceptibility pattern of isolates of *Candida* species to fluconazole show that the most of isolates (20, 31.3%) of were sensitive to antifungal at the concentration of 0.0625 µg/mL. Our results indicated that both non-*albicans* species, *C. glabrata* and *C. tropicalis* isolates were more resistant to fluconazole than *C. albicans*.

Our results described that terbinafine has no valuable effect against different species of *Candida*, especially non-*albicans* species. As shown in Table 7, 30 (46.9%)

Table 3. Age range and sex sampled cases with positive culture (urine)

Age range	Male	Female	Total
< 1 week	8 (38.1%)	1 (4.8%)	9 (42.9%)
1-4 weeks	2 (9.5%)	4 (19.0)	6 (28.5%)
1-6 months	1 (4.8%)	2 (9.5%)	3 (14.2%)
7-12 months	0 (0.0%)	0 (0.0%)	0 (0.0%)
1-6 years	1 (4.8%)	1 (4.8%)	2 (9.6%)
7-12 years	0 (0.0%)	1 (4.8%)	1 (4.8%)
Total	12 (57.1%)	9 (42.9%)	21 (100%)

Table 2. Colony counts of swab cultures

Omeranieme	Colony counts CFU/swab						Total
Organisms –	< 10	11-20	21-30	31-40	41-50	> 50	Total
C. albicans	6 (15.4%)	2 (5.1%)	1 (2.5%)	1 (2.5%)	1 (2.5%)	13 (33.3%)	24 (61.5%)
C. glabrata	4 (10.3%)	1 (2.5%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	5 (12.9%)
C. tropicalis	2 (5.1%)	1 (2.5%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	3 (7.6%)
Candida spp.	2 (5.1%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	5 (12.8%)	7 (18%)
Total	14 (35.9%)	4 (10.3%)	1 (2.5%)	1 (2.5%)	1 (2.5%)	18 (46.1%)	39 (100%)

Table 4. Colony counts of urine cultures

Organisms		Colony counts CFU/mL				
Organisms —	<1000	1001-5000	5001-10000	>10000	Total	
C. albicans	2 (8%)	5 (20%)	1 (4%)	8 (32%)	16 (64%)	
C. glabrata	0 (0.0%)	4 (16%)	0 (0.0%)	1 (4%)	5 (20%)	
C. tropicalis	1 (4%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	1 (4%)	
Candida spp.	0 (0.0%)	3 (12%)	0 (0.0%)	0 (0.0%)	3 (12%)	
Total	3 (12%)	12 (48%)	1 (4%)	9 (36%)	25 (100%)	

Table 5. Sensitivity of isolates of Candida from ICU and NICU to amphotericine B

Organisms	C. albicans	C. glabrata	C. tropicalis	Candida Sp.	Total
Amphotericine B (μg/mL)					
≥ 8	7 (10.9%)	1 (1.5%)	0 (0.0%)	1 (1.5%)	9 (14.1%)
4	10 (15.6%)	4 (6.3%)	1 (1.5%)	6 (9.4%)	21 (32.8%)
2	16 (25%)	1 (1.5%)	0 (0.0%)	2 (3.1%)	19 (29.7%)
1	6 (9.4%)	4 (6.3%)	3 (4.7%)	1 (1.5%)	14 (21.9%)
0.5	1 (1.5%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	1 (1.5%)
MIC 50	2	4	1	4	2
MIC 90	1	1	1	2	1
Total	40 (62.5%)	10 (15.6%)	4 (6.3%)	10 (15.6%)	64 (100%)

Table 6. Sensitivity of isolates of Candida from ICU and NICU to fluconazole

Organisms	C. albicans	C. glabrata	C. tropicalis	Candida Sp.	Total
Fluconazole (µg/mL)					
64	4 (6.3%)	5 (7.8%)	2 (3.1%)	3 (4.7%)	14 (21.9%)
32	5 (7.8%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	5 (7.8%)
16	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)
8	0 (0.0%)	0 (0.0%)	0 (0.0%)	1 (1.5%)	1 (1.5%)
4	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)
2	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)
1	0 (0.0%)	2 (3.1%)	1 (1.5%)	0 (0.0%)	3 (4.7%)
0.5	4 (6.3%)	1 (1.5%)	0 (0.0%)	2 (3.1%)	7 (11.0%)
0.25	8 (12.5%)	0 (0.0%)	1 (1.5%)	0 (0.0%)	9 (14.1%)
0.125	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)
0.0625	17 (26.6%)	1 (1.5%)	0 (0.0%)	2 (3.1%)	20 (31.3%)
0.03125	2 (3.1%)	1 (1.5%)	0 (0.0%)	2 (3.1%)	5 (7.8%)
MIC 50	0.25	64	64	0.5	0.25
MIC 90	0.0625	0.0625	0.25	0.03125	0.0625
Total	40 (62.6%)	10 (15.6%)	4 (6.2%)	10 (15.6%)	64 (100%)

of isolates were resistant to terbinafine at the concentration of \geq 32 $\mu g/mL.$

5. Discussion

Out of several predisposing factors that affect the prevalence of candiduria, long stay in hospitals, especially ICUs and NICUs wards, have an important role. In addition, using several broad spectrum antibiotics, immunosuppressive and corticosteroids

drugs in ICUs and NICUs wards, were increased candiduria. During the several last decades, an increasing in several opportunistic fungal infection was observed. For example, the incidence of candiduria in the United States in 2004 was estimated to be \approx 25 000 cases per year (12). As a results, the presences of candiduria among the hospitalized patients must be taken into account by patients, clinicians and laboratory workers. Several reports show that the

Organisms	C. albicans	C. glabrata	C. tropicalis	Candida Sp.	Total
Terbinafine (µg/mL)					
≥ 32	16 (25%)	5 (7.8%)	3 (4.7%)	6 (9.4%)	30 (46.9%)
16	2 (3.1%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	2 (3.1%)
8	1 (1.5%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	1 (1.5%)
4	2 (3.1%)	1 (1.5%)	0 (0.0%)	0 (0.0%)	3 (4.7%)
2	2 (3.1%)	0 (0.0%)	1 (1.5%)	2 (3.1%)	5 (7.8%)
1	3 (4.7%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	3 (4.7%)
0.5	4 (6.3%)	2 (3.1%)	0 (0.0%)	1 (1.5%)	7 (10.1%)
0.25	10 (15.6%)	2 (3.1%)	0 (0.0%)	1 (1.5%)	13 (20.3%)
MIC 50	4	32	32	32	16
MIC 90	0.25	0.25	2	0.5	0.25
Total	40 (62.5%)	10 (15.6%)	4 (6.3%)	10 (15.6%)	64 (100%)

incidence of candiduria was varied in the different countries. For example, positive urine cultures were reported 2% in Clarke et al (25) study from NICUs patients in Canada. On the other hand, a high rate of Candida colonization, 57.5% and 71.4% was reported in ICU patients by Jain et al (7) and Singla et al (17) in India.

A review of the Iranian literature indicates that the prevalence of candiduria varies considerably in the different province of Iran. For example this rate was reported 32.26% in Qazvin (26), 21.7% in Tehran (27), 16.5% in Ahvaz (5), and 13.8% in Tabriz (28). Our study showed that 21% of samples from hospitalized patients in both ICUs and NICUs were vielded different species of Candida. The frequency of candiduria in our previous study was 5.2% among children attending in children hospital in Ahvaz (10). Although, several studies have shown that candiduria is more frequently in females than males (29-31), we found that candiduria was more common among males (57.1%) than females (42.9%) among children in ICUs and NICUs. In addition, 85.6% of candiduric patients have less than 6 months years old. Our results show that the oral colonization by different species of Candida was found among 34% of hospitalized patients in ICUs and NICUs, so that, 55.9% of them were females and 38.2% have lower than 1 week old. Study of Mendiratta et al shows that the oral mucosal of 77.1% of preterms were colonized by different species of Candida, with comments C. albicans (14). In addition, 60% of colonized children were males and the rest females. Our study shows that 46.1% of sampled cases had heavy colonization (>50 CFU/ swab) of Candida species in their oral cavity. Although the different species of Candida are as oral cavity mycoflora, heavy colonization by Candida among patients in ICUs and NICUs could be an important factor for infection.

In the most reports, C. albicans was detected as prevalent species from UTI, however non-albicans such as C. glabrata and C. tropicalis appeared as an alternative in many studies. The current study revealed that C. albicans (64%) was the most commonly isolated organism in candiduric patients in ICU and NICU followed by C. glabrata (20%), C. tropicalis (4%) and Candida species (12%). Our results agreed with Said et al (32) and Robinson et al (11) who reported that C. albicans was the most common pathogen in NICU patients. However C. tropicalis and C. parapsilosis were as the second agents in the studies of Said et al (32) and Robinson et al (11) which was differed from our results. Several studies have shown that polymicrobial infections occur in 5%-10% of Candida UTIs and C. glabrata appears to be a frequent pathogen with other species (5,8). 14.5% (14.7% from oral cavity and 14.3% from urine samples) polymicrobial growth were observed in our study.

Colony counts more than 1×10³ CFU/mL detected at 88% cases of candiduria whereas 36% of cases were accounted for >10000 CFU/mL. Although several definitions for colony counts of Candida in urine samples were presented, however there is no a standard for it. Some researchers believe that the counts >4000 CFU/mL and <1000 CFU/mL are a marker of infection and normal carriage, respectively. Whereas colony counts between 1000 and 4000 CFU/ mL are shown a borderline case (22). On the other hand, Bukhary (8) have believed that colony counts more than 100000 CFU/mL among the patients without indwelling urinary catheters associated with UTI.

Although, the majority of the reports show that the

most of Candida isolates were susceptible to antifungal drugs, resistance to antifungals (fluconazole) was observed (10, 23, 33). In a study by Mishra et al (23) all C. glabrata, 50% of C. tropicalis and 12.3% of C. albicans isolated from urine samples were resistant to fluconazole. According to Singla et al study (17), resistance to fluconazole detected in 50% of C. glabrata, 27.3% of C. albicans and 18.6% of C. tropicalis isolates were recovered in urine samples of ICU patients. In addition, resistant to fluconazole was detected in 66.7% of urine isolates of Candida (34). Our study indicated that the most tested species of Candida, 45 (70.3%) were sensitive to fluconazole at the concentration of $\leq 8 \mu g/mL$, whereas 14 (21.9%) of isolates had a MIC ≥64 µg/mL and dose dependent (16-32 µg/mL) were only detected at 5 (7.8%) isolates. The MIC 50 and MIC 90 of these isolates were 0.25 and 0.0625 µg/mL, respectively. Similar to previous reports, resistance to fluconazole was found among non-albicans species such as C. glabrata and C. tropicalis (10,35,36).

A single IV dose of amphotericin B can be produced a suitable level for inhibit Candida species in urinary tract and persist in urine for several days (37). In addition, most reports have shown that Candida species were usually sensitive to amphotericin B (10,17,23,38). Susceptibility criteria for amphotericin B cutoff point are as follows, susceptible, MIC≤1 μg/mL, intermediate, MIC 2 μg/mL and resistant, MIC \geq 4 µg/mL (39). In the present study the range of MICs to amphotericin B was from 0.5 to ≥8 µg/mL. C. glabrata and Candida species were less susceptible to amphotericin B and MIC 50 for both species was 4 µg/mL. Terbinafine is basically an anti-dermatophytic agents (40,41), however some researches have shown that terbinafine has an excellent effect against saprophytic fungi, especially, Aspergillus species (42), Candida species from different sources (43,44). Terbinafine susceptibility breakpoints have been defined as follows: ≤8 μg/mL susceptible and $> 8 \mu g/mL$ resistant (45). In our study 50% of isolates were totally sensitive to terbinafine including 75% of C. tropicalis, 60% of Candida species, 50% of C. glabrata, and 45% of C. albicans. In a study by Rathod et al (45), all tested *C. albicans* were sensitive to terbinafine with MICs in the range of 2 to 8 µg/mL. On the other hand the resistant rates of C. albicans was 74.7% for terbinafine in Shi et al study (46).

6. Conclusions

It is concluded that *Candida* colonization has a considerable prevalence among patients hospitalized in NICUs and ICUs in Ahvaz (34% of oral cavity

swabs and 21% of urine samples). As a results, hospitalized patients in critical wards need to major attention for a better control for infections. On the other hand, due to the different susceptibility antifungals results against *Candida* species, the present study showed the need to identify candida recovered from candiduria.

7. Limitations of the study

In the present study only hospitalized neonates and children in ICUs and NICUs were sampled and all obtained data correlated to these groups of patients. In addition, the duration of stay in both wards in hospital was ignored, as a results the sampled patients have different length of hospital stay.

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Authors' Contribution

AZM developed the original idea and the protocol, and edited final manuscript, and is guarantor. ARM contributed to the development of the protocol and edited draft manuscript. MN contributed to the isolation, identification and susceptibility tests and data analysis. MT and SM contributed to present patients for sampling. All authors read and signed final draft.

Conflicts of interest

The authors declared no competing interests.

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References

- Yang YL, Wang AH, Wang CW, Cheng WT, Li SY, Lo HJ. Susceptibilities to amphotericin B and fluconazole of Candida species in Taiwan surveillance of antimicrobial resistance of yeasts 2006. Diagn Microbiol Infect Dis. 2008;61(2):175-80.
- Feng X, Ling B, Yang G, Yu X, Ren D, Yao Z. Prevalence and distribution profiles of Candida parapsilosis, Candida orthopsilosis and Candida metapsilosis responsible for superficial candidiasis in a Chinese university hospital. Mycopathologia. 2012;173(4):229-34.
- Zarrin M, Zarei Mahmoudabadi A. Invasive candidiasis; a review article. Jundishapur J Microbiol.

- 2009;2(1):1-6.
- 4. Tobudic S, Kratzer C, Lassnigg A, Presterl E. Antifungal susceptibility of Candida albicans in biofilms. Mycoses. 2012;55(3):199-204.
- Zarei Mahmoudabadi A, Zarrin M, Ghanatir F, Vazirianzadeh B. Candiduria in hospitalized patients in educational hospitals of Ahvaz. Iran J Microbiol. 2012;4(4):15-24.
- Zarei Mahmoudabadi A, Keradmand AR, Enayatollahi N. Frequency of candiduria in inpatients and outpatients in department of Urology, Golestan Hospital, Ahvaz, Iran. Iran J Kidney Dis. 2009;3(2):114-5.
- Jain M, Dogra V, Mishra B, Thakur A, Loomba PS, Bhargava A. Candiduria in catheterized intensive care unit patients: emerging microbiological trends. Indian J Pathol Microbiol. 2011;54(3):552-5.
- 8. Bukhary ZA. Candiduria: a review of clinical significance and management. Saudi J Kidney Dis Transpl. 2008;19(3):350-60.
- Paul N, Mathai E, Abraham OC, Michael JS, Mathai D. Factors associated with candiduria and related mortality. J Infect. 2007;55:450-5.
- Seifi Z, Azish M, Salehi Z, Zarei Mahmoudabadi A, Shamsizadeh A. Candiduria in children and susceptibility patterns of recovered Candida species to antifungal drugs in Ahvaz. J Nephropathol. 2013;2(2):122-8.
- Robinson JL, Davies HD, Barton M, O'Brien K, Simpson K, Asztalos E, et al. Characteristics and outcome of infants with candiduria in neonatal intensive care-a paediatric investigators collaborative network on infections in Canada (PICNIC) study. BMC Infect Dis. 2009;9(1):183.
- Sobel JD, Fisher JF, Kauffman CA, Newman CA. Candida urinary tract infections-epidemiology. Clin Infect Dis. 2011;52 Suppl 6:S433-6.
- 13. Kauffman CA. Candiduria. Clin Infect Dis. 2005;41 (suppl 6):S371-6.
- Mendiratta D, Rawat V, Thamke D, Chaturvedi P, Chhabra S, Narang P. Candida colonization in preterm babies admitted to neonatal intensive care unit in the rural setting. Indian J Med Microbiol. 2006;24(4):263-7.
- 15. Rathor N, Khillan V, Sarin SK. Nosocomial candiduria in chronic liver disease patients at a hepatobilliary center. Indian J Crit Care Med. 2014;18(4):234-7.
- Fakhri A, Navid M, Seifi Z, Zarei Mahmoudabadi A. The frequency of candiduria in hospitalized patients with depressive syndrome. J Renal Inj Prev. 2014;3(4):97-8.
- 17. Singla N, Gulati N, Kaistha N, Chander J. Candida colonization in urine samples of ICU patients: determination of etiology, antifungal susceptibility

- testing and evaluation of associated risk factors. Mycopathologia. 2012;174(2):149-55.
- 18. Sanclemente G, Marco F, Cervera C, Hoyo I, Colmenero J, Pitart C, et al. Candida norvegensis fungemia in a liver transplant recipient. Rev Iberoam Micol. 2015;32(2):115-7.
- 19. Fekkar A, Dannaoui E, Meyer I, Imbert S, Brossas JY, Uzunov M, et al. Emergence of echinocandin-resistant Candida spp. in a hospital setting: a consequence of 10 years of increasing use of antifungal therapy? Eur J Clin Microbiol Infect Dis. 2014;33(9):1489-96.
- Lin CC, Liu CP, Hsieh FC, Lee CM, Wang WS. Antimicrobial susceptibility and clinical outcomes of Candida parapsilosis bloodstream infections in a tertiary teaching hospital in Northern Taiwan. J Microbiol Immunol Infect. 2014. pii: S1684-1182(14)00163-7.
- 21. Maldonado NA, Cano LE, De Bedout C, Arbelaez CA, Roncancio G, Tabares AM, et al. Association of clinical and demographic factors in invasive candidiasis caused by fluconazole-resistant Candida species: a study in 15 hospitals, Medellin, Colombia 2010-2011. Diagn Microbiol Infect Dis. 2014;79(2):280-6.
- 22. da Costa KR, Ferreira JC, Komesu MC, Candido RC. Candida albicans and Candida tropicalis in oral candidosis: quantitative analysis, exoenzyme activity, and antifungal drug sensitivity. Mycopathologia. 2009;167(2):73-9.
- 23. Mishra M, Agrawal S, Raut S, Kurhade AM, Powar RM. Profile of yeasts isolated from urinary tracts of catheterized patients. J Clin Diagn Res. 2014;8(2):44-6.
- 24. Refrence method for broth dilution abtifungal suceptibility testing of yeasts; approved standard-third edition. M27-A3.28(14).
- Clarke D, Gowrishankar M, Etches P, Lee BE, Robinson JL. Management and outcome of positive urine cultures in a neonatal intensive care unit. J Infect Public Health. 2010;3(4):152-8.
- 26. Ghiasian SA, Aghamirian MR, Eshghi GR. Nosocomial candiduria in critically Ill patients admitted to intensive care units in Qazvin, Iran. Avic J Clin Microbio Infect. 2014;1(2):216-22.
- Tabatabaii SA, Fahimzad A, Shirvani F, Naderi M, Talebian M. Prevalence of nosocomial urinary tract infection in PICU of referral children hospital in Iran. Pak J Med Sci. 2011;27(3):618-21.
- Haggoo SM, Moaddab SR, Sabour S, Varshochi M, editors. Frequency of Candida species isolated from urine cultures in hospitalized patients. The 13th Iranian & The Second International Congress of Microbiology; 2012.
- 29. Artiaga Kobayashi CC, Lisboa Fernandes DF, Miranda KC, de Sousa ED, Rodrigues Silva MR. Candiduria in

- hospital patients: A study prospective. Mycopathologia. 2004;158:49-52.
- Hassaneen AM, Ghonaim RA, Hassanin HM, Salama NA, Elgohary T. Different aspects of candiduria as an important nosocomial infection. Med J Cairo Univ. 2014;82(1):199-204.
- 31. Dalen DM, Zvonar RK, Jessamine PG. An evaluation of the management of asymptomatic catheter-associated bacteriuria and candiduria at the Ottawa hospital. Canadian J Infect Dis Med Microbiol. 2005;16(3):166-70.
- 32. Said RN, Badr AM, Younan MA, Saad MS. Neonatal candiduria: does it jeopardize the outcome of infants at risk in Cairo University Neonatal Intensive Care Units (NICUs)? J Trop Pediatr. 2012;58(5):415-7.
- 33. de Freitas AR, Baeza LC, Faria MG, Dota KF, Godoy Martinez P, Svidzinski TI. Yeasts isolated from nosocomial urinary infections: antifungal susceptibility and biofilm production. Rev Iberoam Micol. 2014;31(2):104-8.
- 34. Yashavanth R, Shiju M, Bhaskar U, Ronald R, Anita K. Candiduria: prevalence and trends in antifungal susceptibility in a tertiary care hospital of Mangalore. J Clin Diagn Res. 2013;7(11):2459-61.
- 35. Yang YL, Li SY, Cheng HH, Lo HJ, Hospitals T. Susceptibilities to amphotericin B and fluconazole of Candida species in TSARY 2002. Diagn Microbiol Infect Dis. 2005;51(3):179-83.
- Zarei Mahmoudabadi A, Zarrin M, Beheshti Fard M. Antifungal susceptibility of Candida species isolated from candidura. Jundishapur J Microbiol. 2013;6(1):24-8.
- 37. Fisher JF, Woeltje K, Espinel-Ingroff A, Stanfield J, DiPiro JT. Efficacy of a single intravenous dose of amphotericin B for Candida urinary tract infections: further favorable experience. Clin Microbiol Infect.

- 2003;9(10):1024-7.
- 38. Ozhak-Baysan B, Ogunc D, Colak D, Ongut G, Donmez L, Vural T, et al. Distribution and antifungal susceptibility of Candida species causing nosocomial candiduria. Med Mycol. 2012;50(5):529-32.
- 39. Ellis D. Amphotericin B: spectrum and resistance. J Antimicrob Chemother. 2002;49(suppl 1):7-10.
- 40. Gupta A, Lynch L, Kogan N, Cooper E. The use of an intermittent terbinafine regimen for the treatment of dermatophyte toenail onychomycosis. J Eur Acad Dermatol Venereol. 2009;23(3):256-62.
- 41. Keyvan P, Leila B, Zahra R, Manuchehr S, Kamiar Z. In vitro activity of six antifungal drugs against clinically important dermatophytes. Jundishapur J Microbiol. 2007;2(4):158-63.
- Zarei Mahmoudabadi A, Seifi Z, Gharaghani M. Lamisil, a potent alternative antifungal drug for otomycosis. Curr Med Mycol. 2015;1(1):18-21.
- Salehei Z, Seifi Z, Zarei Mahmoudabadi A. Sensitivity of vaginal isolates of Candida to eight antifungal drugs isolated from Ahvaz, Iran. Jundishapur J Microbiol. 2012;5(4):574-7.
- Zarei Mahmoudabadi A, Najafyan M, Moghimipour E, Alwanian M, Seifi Z. Lamisil versus clotrimazole in the treatment of vulvovaginal candidiasis. Iran J Microbiol. 2013;5(1):86-90.
- 45. Rathod VS, Raut JS, Mohan Karuppayil S. Antifungal drug susceptibility of Candida albicans isolates from pulmonary tuberculosis patients. Int J Pharm Pharm Sci. 2012;4(Suppl 5):323-6.
- 46. Shi XY, Yang YP, Zhang Y, Li W, Wang JD, Huang WM, et al. Molecular identification and antifungal susceptibility of 186 Candida isolates from vulvovaginal candidiasis in southern China. J Med Microbiol. 2015 doi: 10.1099/jmm.0.000024.

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